

Description

Glycoside-Containing Liposome

5 Technical Field

[0001]

The present invention relates to a liposome
(hereinafter may be referred to as a "liposomal composition")
containing a glycoside which is useful as an anticancer agent.

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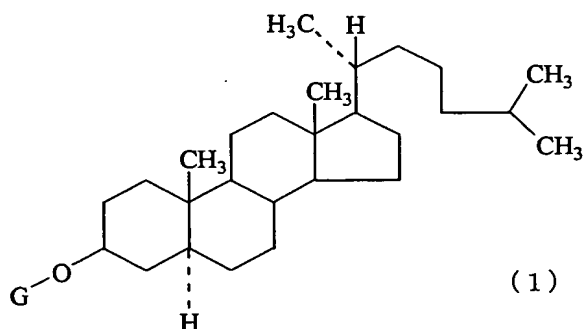
Background Art

[0002]

Among other cholestanol glycosides, the below-described
cholestanol glycoside (1), which is obtained through bonding
15 of a specific sugar chain to cholestanol formed through
saturation of the carbon-carbon double bond of the B ring of
cholesterol, exhibits the effect of inhibiting growth of
cancer cells, and is a compound useful as an anticancer agent
(see Patent Documents 1 and 2).

20 [0003]

[F1]



[0004]

[wherein G represents GlcNAc-Gal-, GlcNAc-Gal-Glc-, Fuc-Gal-, Gal-Glc-, or Gal-].

[0005]

5 A hydrophobic compound such as cholestanol has affinity to cell membranes, and is readily taken into cells. Therefore, a glycoside of such a compound has been considered to be readily taken into various cancer cell lines and to exhibit its effects sufficiently. However, such a glycoside
10 is difficult to use due to its low solubility, and may fail to sufficiently exhibit its effects to some carcinomas.

Patent Document 1: JP-A-11-60592

Patent Document 2: JP-A-2000-191685

15 Disclosure of the Invention

[0006]

An object of the present invention is to provide a drug product which contains a glycoside having antitumor activity, and which enables the glycoside to exhibit its inherent
20 antitumor effects more efficiently.

[0007]

In view of the foregoing, the present inventors have conducted extensive studies on administration forms of a glycoside of a hydrophobic compound (hereinafter may be
25 referred to as a "hydrophobic compound glycoside") having antitumor activity, and as a result have found that when a liposomal composition is prepared from the glycoside together

with a membrane component (at least a phospholipid) and a positive-charge-providing substance, the liposomal composition enables use of the glycoside, which has been difficult to use due to its low solubility, and the

composition enables the glycoside to more efficiently exhibit its inherent antitumor effects; for example, the composition exhibits potent antitumor activity against such a tumor line that scarcely responds to the antitumor effects of the glycoside when administered as such.

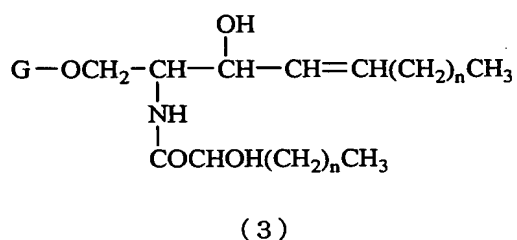
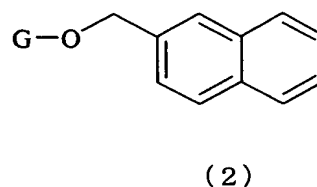
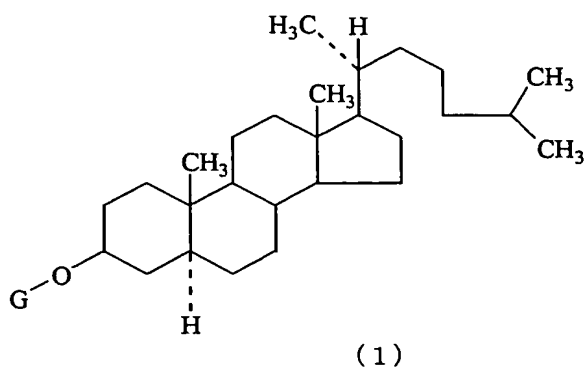
10 [0008]

Accordingly, the present invention provides a liposomal composition containing a glycoside exhibiting antitumor activity, a phospholipid, and a positive-charge-providing substance; the glycoside being composed of GlcNAc-Gal-,
15 GlcNAc-Gal-Glc-, Fuc-Gal-, Gal-Glc-, or Gal- as a sugar moiety, and a hydrophobic compound capable of forming a liposome.

[0009]

The present invention also provides a liposomal
20 composition containing a glycoside selected from among glycosides represented by the following formulas (1) through (3):

[F2]



[0010]

[wherein G represents GlcNAC-Gal-, GlcNAC-Gal-Glc-, Fuc-Gal-, Gal-Glc-, or Gal-, and n represents an integer of 12 to 26];

5 a phospholipid; and a positive-charge-providing substance.

[0011]

The present invention also provides an anticancer agent containing the liposomal composition.

[0012]

10 According to the present invention, the solubility of a glycoside having antitumor activity is enhanced, and there can be provided a drug product which enables the glycoside to more efficiently exhibit its inherent antitumor effects.

Particularly, a drug product containing cholestanol glycoside

15 (1) exhibits potent antitumor activity against such a cancer

cell line that scarcely responds to the antitumor effects of the glycoside when administered as such; i.e., the drug product can maximize the antitumor effects of the compound.

5 Brief Description of the Drawings

[0013]

[Fig. 1] Fig. 1 shows the cancer cell proliferation inhibitory effect of GlcNAc β 1,4GalChol-containing liposomes. In Fig. 1, "-Lipo" represents liposome.

10 [Fig. 2] Fig. 2 shows the cancer cell proliferation inhibitory effect of Fuc α 1,3GalChol-containing liposomes. In Fig. 2, "-Lipo" represents liposome.

[Fig. 3] Fig. 3 shows the cancer cell proliferation inhibitory effect of Gal β Chol-containing liposomes. In Fig.

15 3, "-Lipo" represents liposome.

[Fig. 4] Fig. 4 shows the cancer cell proliferation inhibitory effect of GlcNAc β 1,3Gal β 1,1NM-containing liposomes or GlcNAc β 1,3Gal β 1,4Glc β 1,1Ceramide-containing liposomes. In Fig. 4, "-Lipo" represents liposome.

20 [Fig. 5] Fig. 5 shows the peritoneal dissemination inhibitory effect of Fuc α 1,3GalChol-containing liposomes having different particle sizes.

[Fig. 6] Fig. 6 shows the peritoneal dissemination inhibitory effect of GlcNAc β 1,4GalChol-containing liposomes, particularly shows the results of counting the number of

25 tumors formed.

[Fig. 7] Fig. 7 shows the peritoneal dissemination

inhibitory effect of GlcNAc β 1,4GalChol-containing liposomes, particularly shows the results of measuring tumor weight (g). [Fig. 8] Fig. 8 shows comparison in peritoneal dissemination inhibitory effect between administration of FucGalChol-containing liposomes and administration of FucGalChol alone, particularly shows the results of counting the number of tumors formed.

Best Mode for Carrying Out the Invention

[0014]

10 The glycoside contained in the liposomal composition of the present invention includes, as a sugar moiety, GlcNAc-Gal-, GlcNAc-Gal-Glc-, Fuc-Gal-, Gal-Glc-, or Gal-, includes, as an aglycon, a hydrophobic compound capable of forming liposomes, and has antitumor activity.

15 Examples of the hydrophobic compound capable of forming liposomes include biocomponents such as cholesterol, ceramide, hydrophobic amino acids, fatty acids (e.g., oleic acid, linoleic acid, and linolenic acid), and fat-soluble vitamins; compounds having an aromatic ring, such as naphthalene derivatives (e.g., naphthalene methanol); and cholesterol derivatives (e.g., cholestanol). Of these, ceramide, naphthalene derivatives, and cholesterol derivatives are preferred, with cholestanol, which is formed through saturation of the carbon-carbon double bond of the B ring of
20 cholesterol, being particularly preferred.

[0015]

When the sugar moiety is GlcNAc-Gal-, GlcNAc β 1,3-Gal β -

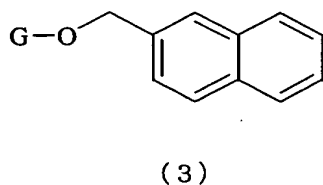
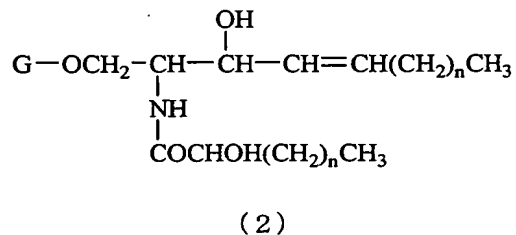
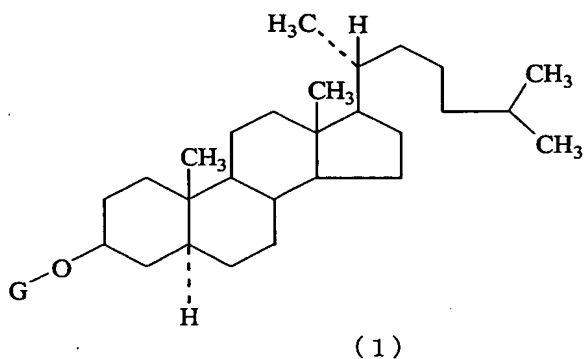
or GlcNAc β 1,4-Gal β - is preferred. When the sugar moiety is GlcNAc-Gal-Glc-, GlcNAc β 1,3-Gal β 1,4-Glc- is preferred. When the sugar moiety is Fuc-Gal-, Fuc α 1,3Gal- is preferred. When the sugar moiety is Gal-Glc-, Gal β 1,4Glc β - is preferred.

5 When the sugar moiety is Gal-, Gal β - is preferred.

[0016]

Preferred examples of the glycoside include the following cholestanol glycoside (1), naphthalene methanol glycoside (2), and ceramide glycoside (3):

10 [F3]



[0017]

[wherein G represents GlcNAc-Gal-, GlcNAc-Gal-Glc-, Fuc-Gal-, Gal-Glc-, or Gal-, and n represents an integer of 12 to 26].

15 [0018]

Of these, cholestanol glycoside (1) is particularly

preferred, from the viewpoint of anticancer effects.

Preferred examples of the glycoside represented by formula

(1) include the following compounds:

(1) G = GlcNAc-Gal-:

5 3- β -cholestanyl 3-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- β -D-galactopyranoside, and

 3- β -cholestanyl 4-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- β -D-galactopyranoside;

(2) G = GlcNAc-Gal-Glc-:

10 3- β -cholestanyl 4-O-{3-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- β -D-galactopyranosyl}- β -D-glucopyranoside;

(3) G = Fuc-Gal-:

 3- β -cholestanyl 3-O-(α -L-fucopyranosyl)- β -D-galactopyranoside;

15 (4) G = Gal-Glc-:

 3- β -cholestanyl 4-O-(β -D-galactopyranosyl)- β -D-glucopyranoside; and

(5) G = Gal-:

 3- β -cholestanyl β -D-galactopyranoside.

20 [0019]

The glycoside employed in the present invention can be extracted from a living organism, or produced through known chemical reaction. Alternatively, the glycoside can be obtained through combination of such extraction and reaction.

25 For example, the aforementioned cholestanol glycoside (1) can be produced through the method described in JP-A-11-60592 or JP-A-2000-191685; the naphthalene methanol glycoside (2) can

be produced through the method described in J. Biol. Chem.
272 (41) 25608, 1997; and the ceramide glycoside (3) can be
extracted from, for example, human erythrocytes, liver,
gastrointestinal mucosa, or meconium.

5 [0020]

The glycoside has antitumor activity. As used herein,
the expression "the glycoside has antitumor activity" refers
to the case where, in vivo, the glycoside exhibits antitumor
effects against at least one carcinoma, or where, in vitro,
10 the glycoside has activity to inhibit or suppress growth of
at least one cancer cell line.

[0021]

The amount of the glycoside contained in the liposomal
composition of the present invention is 0.3 to 2.0 mol,
15 preferably 0.8 to 1.5 mol, on the basis of 1 mol of a
phospholipid.

[0022]

The liposomal composition of the present invention
contains at least a phospholipid as a membrane component.

20 Examples of the phospholipid include
phosphatidylcholines such as dilauroylphosphatidylcholine,
dimyristoylphosphatidylcholine,
dipalmitoylphosphatidylcholine, distearoylphosphatidylcholine,
dioleoylphosphatidylcholine, dilinoleoylphosphatidylcholine,
25 myristoylpalmitoylphosphatidylcholine,
myristoylstearylphosphatidylcholine, and
palmitoylarachidoylphosphatidylcholine;

phosphatidylethanolamine; phosphatidylserine;
phosphatidylinositol; and phosphatidic acid. The
phospholipid may be a naturally occurring product, or may be
obtained through semisynthesis or total synthesis. The
5 phospholipid may be a processed phospholipid such as a
hydrogenated phospholipid. These phospholipids may be
employed singly or in combination of two or more species.

Of the aforementioned phospholipids,
phosphatidylcholines are preferably employed, and 1 α -
10 dipalmitoylphosphatidylcholine (DPPC) is particularly
preferably employed, since electrically neutral, stable
liposomes can be formed.

[0023]

The positive-charge-providing substance is added for
15 positively charging the surface of a lipid membrane. When
the surface of liposomes is positively charged, the liposomes
are expected to come into natural contact with cells having
negatively charged membrane surfaces.

[0024]

20 Examples of the positive-charge-providing substance
include aliphatic amines such as stearylamine and oleylamine;
and aromatic amines such as fluoreneethylamine. Of these,
aliphatic amines are preferred, and stearylamine is
particularly preferably employed.

25 The amount of the positive-charge-providing substance
contained in the liposomal composition is 0.04 to 0.15 mol,
preferably 0.1 to 0.15 mol, on the basis of 1 mol of the

phospholipid.

[0025]

If desired, the liposomal composition of the present invention may contain, in addition to the aforementioned components, a membrane structure stabilizer such as cholesterol, a fatty acid, or diacetyl phosphate.

[0026]

The aqueous solution employed for dispersing the membrane component is preferably water, saline, a buffer, an aqueous solution of a sugar, or a mixture thereof. The buffer to be employed is preferably an organic or inorganic buffer which has buffering action in the vicinity of the hydrogen ion concentration of body fluids. For example, a phosphate buffer can be employed.

[0027]

No particular limitation is imposed on the method for preparing the liposomal composition of the present invention, and the composition can be prepared through a customary method. For example, the liposomal composition can be prepared through the method described in JP-A-57-82310, JP-A-60-12127, JP-A-60-58915, JP-A-1-117824, JP-A-1-167218, JP-A-4-29925, or JP-A-9-87168, the method described in Methods of Biochemical Analysis (1988) 33, p 337, or the method described in "Liposome" (Nankodo).

[0028]

Next will be described preparation of the liposomal composition of the present invention through the method

described in Japanese Patent Application Laid-Open (*kokai*) No. 9-87168.

[0029]

Firstly, an organic solvent and water are added to and
5 mixed with a glycoside, a phospholipid, and a positive-charge-providing substance, and subsequently the organic solvent is completely removed by means of a rotary evaporator or a similar apparatus, followed by removal of the water. In this case, the mixing proportions of the membrane component,
10 the positive-charge-providing substance, and the glycoside may be, for example, 52 : 8 : 20 (by mole). However, so long as the mixing proportions fall within a range nearly equal to the above range, particular problems do not arise. When the mixing proportion of the glycoside is low, if desired, a
15 membrane structure stabilizer such as cholesterol may be added. However, when the mixing proportion of the glycoside is high, addition of such a membrane structure stabilizer is not necessarily required.

[0030]

20 No particular limitation is imposed on the organic solvent to be employed, so long as it is a volatile organic solvent which is insoluble in water. Examples of the organic solvent which may be employed include chloroform, chloromethane, benzene, and hexane. In consideration of
25 solubility, an organic solvent having relatively high polarity (e.g., ethanol or methanol) may be appropriately added to such a water-insoluble solvent, and the thus-

prepared organic solvent mixture may be employed. No particular limitation is imposed on the mixing proportions of the organic solvent mixture and water, so long as a uniform solvent mixture is obtained.

5 [0031]

In the case where water is added for preparation of the liposomal composition, removal of the water is generally carried out through freeze-drying. However, removal of the water is not necessarily performed through freeze-drying, and may be performed through drying in a reduced-pressure desiccator. After removal of the water, the aforementioned aqueous solution for dispersion is added, followed by impregnation by means of, for example, a Vortex mixer, to thereby form the liposomal composition.

15 [0032]

The particle size of the liposomal composition of the present invention is preferably 10 μm or less, more preferably 3 μm or less, from the viewpoint of tumor-suppressive effect.

20 Liposomes having a uniform particle size can be prepared through, for example, ultrasonic treatment, extrusion treatment by use of a porous membrane filter, treatment by use of a high-pressure injection emulsifier, or combination of such treatments. Smaller liposome particles can be prepared by, for example, performing ultrasonic treatment for a long period of time.

[0033]

The thus-prepared liposomal composition of the present invention exhibits very excellent effect of inhibiting growth of cancer cells, as described below in Examples.

Particularly when the cholestanol glycoside (1) is employed, the liposomal composition exhibits potent antitumor activity against such a cancer cell line that scarcely responds to the antitumor effects of the glycoside when administered as such.

In general, liposomes are vesicles formed of phospholipid bilayer membranes, and liposomes encapsulating a drug are prepared for the purpose of bringing the drug into cells when they are dispersed and fused with the cell membranes.

Accordingly, the properties of the drug have been considered unchanged through formation of liposomes from the drug.

Therefore, it is quite surprising that the antitumor activity of the cholestanol glycoside is drastically enhanced through formation of a liposomal composition from the cholestanol glycoside.

Thus, a drug product containing the liposomal composition is useful as an anticancer agent which enables the glycoside to more efficiently exhibit its inherent antitumor effects.

[0034]

The product form of the anticancer agent of the present invention can be appropriately determined depending on the treatment site or the therapeutic purpose. So long as an additive which impedes the stability of the form of the liposomal composition is not employed, the anticancer agent

can be prepared through a known preparation method into, for example, a peroral product, an injection, a suppository, an ointment, or a patch.

[0035]

5 A peroral solid product (e.g., a tablet, a coated tablet, a granule, a powder, or a capsule) can be prepared by adding, to the liposomal composition of the present invention, an excipient and, if desired, other additives such as a binder, a disintegrating agent, a lubricant, a coloring agent, 10 a flavoring agent, and a deodorant, followed by customary processing. The additive to be employed may be an additive which is generally used in the art. Examples of the excipient include lactose, sucrose, sodium chloride, glucose, starch, calcium carbonate, kaolin, microcrystalline cellulose, 15 and silicic acid. Examples of the binder include water, ethanol, propanol, simple syrup, liquid glucose, liquid starch, liquid gelatin, carboxymethyl cellulose, hydroxypropyl cellulose, hydroxypropyl starch, methyl cellulose, ethyl cellulose, shellac, calcium phosphate, and 20 polyvinyl pyrrolidone. Examples of the disintegrating agent include dry starch, sodium alginate, agar powder, sodium hydrogencarbonate, calcium carbonate, sodium lauryl sulfate, monoglyceride stearate, and lactose. Examples of the lubricant include purified talc, stearic acid salts, borax, 25 and polyethylene glycol. Examples of the flavoring agent include sucrose, orange peel, citric acid, and tartaric acid.

[0036]

A peroral liquid product (a peroral solution, a syrup, or an elixir) can be prepared by adding, to the liposomal composition of the present invention, a flavoring agent, a buffer, a stabilizer, a deodorant, or the like, followed by customary processing. The flavoring agent to be employed for this preparation may be the aforementioned one. Examples of the buffer include sodium citrate. Examples of the stabilizer include tragacanth, gum arabi, and gelatin.

[0037]

An injection (e.g., a subcutaneous injection, an intramuscular injection, or an intravenous injection) can be prepared by adding, to the liposomal composition of the present invention, a pH-adjusting agent, a buffer, a stabilizer, an isotonicity-imparting agent, a local anesthetic agent, or the like, followed by customary processing. Examples of the pH-adjusting agent and the buffer employed for this preparation include sodium citrate, sodium acetate, and sodium phosphate. Examples of the stabilizer include sodium pyrosulfite, EDTA, thioglycolic acid, and thiolactic acid. Examples of the local anesthetic agent include procaine hydrochloride and lidocaine hydrochloride. Examples of the isotonicity-imparting agent include sodium chloride and glucose.

[0038]

A suppository can be prepared by adding, to the liposomal composition of the present invention, a known carrier for drug preparation, such as polyethylene glycol,

lanolin, cocoa butter, or fatty acid triglyceride, and, if desired, a surfactant such as Tween (registered trademark), followed by customary processing.

[0039]

5 An ointment can be prepared by mixing the liposomal composition of the present invention with, if desired, a commonly employed additive such as a base, a stabilizer, a humectant, or a preservative through a customary technique. Examples of the base include liquid paraffin, white
10 petrolatum, white beeswax, octyldodecyl alcohol, and paraffin. Examples of the preservative include methyl p-hydroxybenzoate, ethyl p-hydroxybenzoate, and propyl p-hydroxybenzoate.

[0040]

A patch can be prepared in a routine manner by applying,
15 to a commonly employed support, the aforementioned ointment, a cream, a gel, a paste, or the like. Examples of appropriate supports include woven fabrics and non-woven fabrics formed of cotton, staple fiber, and chemical fiber; and films and foam sheets formed of soft vinyl chloride,
20 polyethylene, and polyurethane.

[0041]

The dose of the anticancer agent of the present invention varies depending on the symptom, body weight, age, sex, etc. of a patient in need thereof. The daily dose of
25 the anticancer agent for an adult is typically about 0.01 to about 200 mg/Kg, preferably 0.1 to 50 mg/Kg, as reduced to the cholestanol glycoside (1). Preferably, the daily dose is

administered once a day, or in a divided manner (2 to 4 times a day).

Examples

[0042]

5 The present invention will next be described in more detail by way of Examples.

Example 1 Formation of liposomes from cholestanol glycoside

There were employed, as cholestanol glycosides, "GlcNAc β 1,4GalChol"; i.e., a cholestanol compound of formula
10 (1) in which G is GlcNAc β 1,4Gal, "Fuc α 1,3GalChol"; i.e., a cholestanol compound of formula (1) in which G is Fuc α 1,3Gal, and "Gal β Chol"; i.e., a cholestanol compound of formula (1) in which G is Gal β . For comparison, "Chol"; i.e., a cholestanol compound of formula (1) in which G is H was
15 employed. A 20 μ mol/mL solution of each of these compounds (dissolved in chloroform/methanol = 5/1 (v/v)) was employed as a starting material.

1 α -Dipalmitoylphosphatidylcholine, stearylamine, and each of the aforementioned cholestanol compounds were mixed
20 in proportions of 52/8/20 (by mole) so as to attain a total amount of 700 μ L, and subsequently an organic solvent (chloroform/methanol = 2/1 (v/v)) (300 μ L) and distilled water (1 mL) were added to and mixed with the resultant mixture. Thereafter, the organic solvent was completely
25 removed by means of a rotary evaporator, and the resultant product was subjected to freeze-drying, to thereby completely remove the water. The freeze-dried product was dissolved in

PBS (1 mL), followed by ultrasonic treatment (15 W, 15 minutes), to thereby form liposomes having a uniform particle size of about 2 to about 4 μm . The thus-formed liposomes were employed in Examples 2 and 3.

5 [0043]

Example 2 Formation of liposomes from naphthalene methanol glycoside

There was employed, as a naphthalene methanol glycoside, "GlcNac β 1,4Gal β 1,1NM" (NM: naphthalene methanol); i.e., a
10 compound of formula (2) in which G is GlcNac β 1,4-Gal β 1,1-, and liposomes were formed from this naphthalene methanol glycoside under conditions similar to those for formation of the cholestanol glycoside liposomes.

[0044]

15 Example 3 Formation of liposomes from ceramide glycoside

There was employed, as a ceramide glycoside, "GlcNac β 1,3Gal β 1,4Glc β 1,1Ceramide"; i.e., a compound of formula (3) in which G is GlcNac β 1,3-Gal β 1,4-Glc-, and liposomes were formed from this ceramide glycoside in a
20 manner similar to that of formation of the cholestanol glycoside liposomes. In the case of formation of ceramide glycoside liposomes, cholesterol was added as a stabilizer.

[0045]

Example 4 Cell proliferation inhibitory effect of
25 cholestanol glycoside liposomes

A cultured cancer cell line (colon26 wild) was inoculated into a 96-well plate (1×10^4 cells/100 μL /well),

and subsequently a cholestanol glycoside (GlcNAc β 1,4GalChol, Fuc α 1,3GalChol, Gal β Chol, or Chol) or each type of the cholestanol glycoside liposomes formed in Example 1 was added to the 96-well plate, followed by incubation at 37°C for three days. Thereafter, the MTT assay was performed, and the number of live cells was determined. The cell proliferation inhibition rate was obtained by use of the following formula.

[0046]

Cell proliferation inhibition rate(CPI rate) (%) = (1 - OD of treated cells/OD of non-treated cells) \times 100

[0047]

As a result, Fuc α 1,3GalChol, which scarcely exhibits cell proliferation inhibitory effect when employed alone, exhibited remarkable cell proliferation inhibitory effect through formation of liposomes therefrom. Meanwhile, GlcNAc β 1,4GalChol, which has conventionally been shown to have cell proliferation inhibitory effect, was found to exhibit further enhanced cell proliferation inhibitory effect through formation of liposomes therefrom (Figs. 1, 2, and 3).

[0048]

Example 5 Cell proliferation inhibitory effect of naphthalene methanol glycoside liposomes and ceramide glycoside liposomes

A cultured cancer cell line (colo201) was inoculated into a 96-well plate (1×10^4 cells/100 μ L/well), and subsequently the naphthalene methanol glycoside liposomes formed in Example 2 or the ceramide glycoside liposomes

formed in Example 3 were added to the 96-well plate, followed by incubation at 37°C for three days. Thereafter, the MTT assay was performed, and the number of live cells was determined. The cell proliferation inhibition rate was obtained in a manner similar to that of Example 4.

As a result, GlcNAc β 1,3Gal β 1NM or GlcNAc β 1,3Gal β 1,4Glc β 1,1Ceramide, which scarcely exhibits cell proliferation inhibitory effect when employed alone due to its insolubility, was found to exhibit remarkable cell proliferation inhibitory effect through formation of liposomes therefrom (Fig. 4).

[0049]

Example 6 Peritoneal dissemination inhibitory effect of cholestanol glycoside liposomes

(1) Effect of the particle size of Fuc α 1,3GalChol liposomes

Balb/c mice (8 weeks old, female) were employed. A cancer cell line (colon26 wild, 5×10^4 cells/200 μ L) was intraperitoneally administered to each of the animals. Subsequently, 12 hours, 24 hours, and 48 hours after the cancer cell line administration, Fuc α 1,3GalChol liposomes (2 μ mol/100 μ L) which had been prepared through ultrasonic treatment (15 W \times 5, 10, or 15 minutes) were intraperitoneally administered. Ten days later, the number of tumors at the mesentery was counted. Liposomes having a smaller particle size, which were prepared through prolonged ultrasonic treatment, were found to exhibit higher peritoneal dissemination inhibitory effect (Fig. 5).

[0050]

(2) Peritoneal dissemination inhibitory effect of
GlcNAc β 1,4GalChol liposomes

Balb/c mice (8 weeks old, female) were employed. A
5 cancer cell line (colon26 wild, 5×10^4 cells/200 μ L) was
intraperitoneally administered to each of the animals.
Subsequently, 24 hours and 48 hours after the cancer cell
line administration, GlcNAc β 1,4GalChol liposomes (2 μ mol/100
10 μ L) which had been prepared through ultrasonic treatment (15
W, 15 minutes) were intraperitoneally administered. Ten days
after the liposome administration, the number of tumors at
the mesentery was counted, and 21 days after the liposome
administration, the weight of tumors formed at the greater
omentum and the mesentery was measured. As a result, the
15 administered GlcNAc β 1,4GalChol liposomes were found to
exhibit excellent peritoneal dissemination inhibitory effect
(Figs. 6 and 7).

[0051]

(3) Comparison in peritoneal dissemination inhibitory effect
20 between administration of FucGalChol liposomes and
administration of FucGalChol alone

Balb/c mice (8 weeks old, female) were employed. A
cancer cell line (colon26 wild, 5×10^4 cells/200 μ L) was
intraperitoneally administered to each of the animals.
25 Subsequently, 0, 24, and 48 hours after the cancer cell line
administration, FucGalChol liposomes which had been prepared
through ultrasonic treatment (15 W, 15 minutes), or

FucGalChol was intraperitoneally administered ($2\text{ }\mu\text{mol}/100\text{ }\mu\text{L}$). Ten days later, the amount of tumors formed at the mesentery was counted.

As a result, tumor formation was found to be strongly
5 inhibited in an FucGalChol liposome administration group, as compared with the case of a non-treated group or an FucGalChol administration group (Fig. 8).